mapped, is not a hotspot of chromosomal breakpoints seen in canine tumours. The hotspots that have been found in the dog genome so far, include chromosomes 1, 19 and 25 which are preferentially involved in chromosomal fusions<sup>4</sup>. The X-chromosome of the dog, in contrast, is frequently affected by structural aberrations. Therefore, in contrast to humans, the activation of *HMGA1* as a result of chromosomal translocations does not seem to play a considerable role in canine tumours. This may be due to the fact that the corresponding changes are not able to induce benign tumours in the dog or to stimulate their growth. Alternatively, there may be factors favouring the occurrence of the structural changes in humans which are lacking in dogs.

No homology has been found between human chromosome 6, to which the HMGA1 is mapped, and canine chromosome 23, rather human chromosome 6 shares homology with canine chromosome 22 and  $8^5$ . In our FISH studies no metaphase shows signals on these dog chromosomes.

## References

- 1 Kazmierczak B. et al. (1996) J Natl Cancer Inst 88, 1234–6.
- 2 Dal Cin P. et al. (1999) Genes Chromosomes Cancer 24, 286-9.
- 3 Murua Escobar H. et al. (2001) Cytogenet Cell Genet 94, 194–5.
- 4 Reimann N. et al. (1999) J Natl Cancer Inst 91, 1688-9.
- 5 Yang F. et al. (1999) Genomics 62, 189-202.

Correspondence: Dr J. Bullerdiek (bullerd@uni-bremen.de)

## Contribution of a new set of canine microsatellites to the knowledge of the canine genetic map

## D. Parra\*, G. Dolf<sup>†</sup> and S. Dunner\*

\*Laboratorio de Genética Molecular, Facultad de Veterinaria, Universidad Complutense de Madrid, 28040 Madrid, Spain. †Institute of Animal Genetics, University of Berne, Switzerland Accepted 28 October 2002

Source/description: Canis familiaris DNA was digested with Sau3AI and fragments ranging 700–1000 bp in length were ligated into the BamHI site of a pre-digested and de-phosphorylated pUC18 vector (Amersham Pharmacia Biotech, Amersham, UK) and transformed into Escherichia coli  $SURE^{\otimes}$  cells (Stratagene, La Jolla, CA, USA). A total of 1200 recombinant clones were pooled in groups of 10 and the pools were spotted onto a nylon membrane, screened with digoxigenin-labelled probes  $(TG)_{10}$  and  $(AAAT)_7$ , and the positive pools were screened individually to isolate the final positive clones. Ten positive clones were sequenced in an ABI-310 sequencer (Applied Biosystems, Foster City, CA, USA) from which six

able 1 Features of the microsatellites, including heterozygosis (H), polymorphic information content (PIC)<sup>2</sup>, effective number of alleles (ENA)<sup>3</sup>, number of informative meiosis (NIM), linkage Chromoocation CFA16 CFA10 CFA5 some CFA1 linkage group Assigned L28/33 L18/33 L19 126 NIM (%) 78 (36) 110 (51) 136 (64) (32)69 ENA 5.8 2.1 3.1 63.5 6.99 80.5 48.1 0 74 44 51.4 67 82. ェ 0 Alleles 3 2  $\infty$ Annealing (°C) 28 28 20 28 63 28 Size range 135-151 103-121 225-241 (dq) 86 154 (TG)<sub>19</sub>(AG)<sub>14</sub> (CT)<sub>13</sub>(GT)<sub>9</sub> (AAAT)<sub>12</sub> (GT)<sub>20</sub> (AC)<sub>14</sub> (CT)<sub>6</sub> unit ACTCAAAAGCCATCTTGTCACA GGTCGACTCTAGAGGATCTGG AAGAAGCTGCAGGAGCTACG GGAGCTCTCTGCGCAGTATG SCGCTTTCTCCAAGTAACAG SAGCAGCAGCCTGGACTAC CATTCCTTTTCATGGTTAAT Primer sequences  $(5' \rightarrow 3')$ **ATCTGTAGTAAAGCTGAC** CATGGAGCCTGCTTCTCC group and chromosomal location. 0 AY059379 AF210623 Accession AF210622 AF210620 AF448484 AF21062 JCMCF117 JCMCF12 JCMCF40 JCMCF54 JCMCF96 JCMCF71 Locus

true-positive clones were obtained (60%) and are detailed in Table 1. Sequences were checked against public database using the BLAST¹ facility and it was observed that clones UCMCF71 from nt591 to nt696 and UCMCF117 in positions 1–49 showed high levels of sequence identity (>80%) with the C. familiaris t-RNA family derived short interspersed nucleotide element (SINE) described by Bentolila  $et\ al.^2$  (GenBank AJ239530–49). UCMCF54 contains not only a SINE at nucleotide positions 9–52 (identity = 98% with AJ239533) but also a C. familiaris LINE-1 element ORF-2 mRNA (GenBank AB012223) in positions 194–328 (82% of sequence identity). Primer pairs for PCR amplification of the microsatellite of the positive clones were designed using the program PRIMER 0.5 (GCG Software Package, University of Wisconsin, WI, USA).

*PCR conditions:* Polymerase chain reaction was performed in a total volume of 10 μl of the following mixture: 10 ng of canine genomic DNA, 5 pmol of each primer, 200 μm of each dNTP, 2 mm MgCl<sub>2</sub>, 1X PCR Buffer [75 mm Tris–HCl (pH 9.0), 50 mm KCl, 20 mm (NH4)<sub>2</sub>SO<sub>4</sub>, 0.001% BSA] and 0.3 U of thermostable DNA polymerase (Biotools, Madrid, Spain, 28080, E). The cycling programme in a PTC-200<sup>TM</sup> Programmable Thermal Controller (MJ Research Inc., Watertown, MA, USA) included initial denaturation during 5 min at 95 °C, 30 cycles of 50 s at 95 °C, 50 s at 50–63 °C, 50 s at 72 °C and final extension at 72 °C for 25 min. PCR products were analysed by capillary electrophoresis in an ABI-310 sequencer (Applied Biosystems).

Polymorphism: Polymorphism was revealed by genotyping 10 unrelated individuals of each of the following breeds: Beagle, English Pointer, English Setter, Epagneul Breton and German Shepherd. The observed alleles were named according to their size in base pairs as estimated with the Genescan® 2.1 software (Applied Biosystems). Markers UCMCF54 and UCMCF117 did not show any polymorphism, but in the rest of markers, heterozygosity (H) and polymorphic information content³ (PIC) was found to range between 51.5–82.7% and 48.1–80.5%, respectively (Table 1). The effective number of alleles⁴ observed ranged between 1 and 5.8.

Mendelian inheritance: Codominant segregation was observed in the Reference Families of the Dogmap Panel<sup>5</sup> including 212 meiosis, where no mutation events or null alleles were observed and the informative meiosis ranged between 69 (32%) and 136 (64%).

Chromosomal location: Linkage analysis was carried out with the software package  $_{\mbox{\footnotesize CRI-MAP}}\mbox{}2.4^6.$  Markers were assigned to previously described linkage groups  $^7$  and anchored into chromosomes by using the 'twopoint' option, and arranged with the 'build' option. Linkage groups and chromosomal locations of the new loci are shown in Table 1 and depicted in Fig. 1.

Acknowledgements: This work has been funded by the Federación Espanola de Caza. We wish to thank the DogMap Consortium for providing the reference panel.

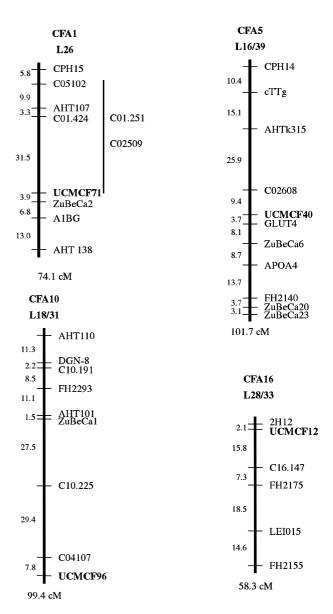


Figure 1 Microsatellite chromosomal location. Distances are given in Kosambi cM. The linkage groups are displayed as sex-averaged maps.

## References

- 1 Altschul S.F. et al. (1990) J Mol Biol 215, 403-10.
- 2 Bentolila S. et al. (1999) Mamm Genome 10, 699-705.
- 3 Botstein D. et al. (1980) Am J Hum Genet 32, 314-31.
- 4 Kimura M. & Crow, J. F. (1964) Genetics 49, 725-38.
- 5 Lingaas F. et al. (1997) Mamm Genome 8, 218–21.
- 6 Lander E. & Green P. (1987) *Proc Nat Acad Sci USA* **84**, 2363–7.
- 7 Lingaas F. et al. (2001) J Anim Breed Genet 118, 3–19.

Correspondence: David Parra (dparra@vet.ucm.es)